# Acute Exposure to Imidacloprid Impairs Walking Behavior of *Eocanthecona furcellata* (Hemiptera: Pentatomidae)<sup>1</sup>

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**Abstract** Imidacloprid, one of the neonicotinoid pesticides, is widely used in various types of farmlands globally, causing negative effects on beneficial insects. The stink bug *Eocanthecona furcellata* (Wolff) (Hemiptera: Pentatomidae) is an effective generalist predator that provides natural control of Lepidoptera, Coleoptera, and other insect pests. Walking behavior is crucial for these predators; however, the effects of imidacloprid on their walking, and thus searching, behavior remain unknown. We measured walking behavior by using a locomotion compensator to assess the effects of imidacloprid. Acute exposure to imidacloprid significantly reduced the mean walking speed of nymphs (30.6–51.9% depending on instar), adult females (52.0%), and adult males (40.5%). Walking time also was reduced by exposure to imidacloprid in nymphs (27.1–40.5%), adult females (48.2%), and adult males (36.0%), whereas walking distance was reduced in nymphs (8.4–19.8%), adult females (25.2%), and adult males (15.0%). These results reveal that imidacloprid impairs the walking behavior of stink bugs, emphasizing the need to consider the negative effects of neonicotinoid pesticides on the pest control efficacy of natural enemy insects.

Key Words imidacloprid, walking behavior, *Eocanthecona furcellata* 

Neonicotinoid pesticides were introduced into the market in the 1990s and rapidly became the fastest growing and most widely used class of pesticides globally (Jeschke et al. 2011; Simon-Delso et al. 2015). They can easily enter the soil and be absorbed by plants, and they are extensively used as seed dressings or foliar sprays in agriculture (Blacquière et al. 2012; Bonmatin et al. 2015; Godfray et al. 2015; Mitchell et al. 2017; Robin and Andreas 2003). Neonicotinoids disrupt the neural activity of insects by binding to nicotinic acetylcholine receptors (Elbert et al. 2008; Matsuda et al. 2001). Furthermore, due to their water solubility and persistence in the environment, neonicotinoids are ingested by herbivorous insects, subsequently exposing predatory insects (Blacquière et al. 2012). Imidacloprid, one of the most common neonicotinoid pesticides, has been restricted for outdoor use in the European Union,

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as have thiamethoxam and clothianidin (Environmental Protection Agency 2015); however, it remains widely used in agricultural production worldwide. Although imidacloprid effectively controls pests, it also adversely affects beneficial insects. Previous studies have shown that imidacloprid can affect many aspects of insect behavior, especially in honey bees and bumblebees, which are important pollinators (Desneux et al. 2007; Kenna et al. 2019; Mengoni Goñalons and Farina 2018; Mustard et al. 2020; Smith et al. 2020; Stanley et al. 2015).

The stink bug *Eocanthecona furcellata* (Wolff) (Hemiptera: Pentatomidae) is regarded as an important generalist predator of pest insects and is widely distributed in China (Yunnan, Sichuan, Hainan, Fujian, Guangdong, Guangxi), India, Sri Lanka, Myanmar, Malaysia, the Philippines, Indonesia, Japan, and Korea (He et al. 2013; Keerthi et al. 2020; Rider and Zheng 2002; Roca-Cusachs et al. 2020; Shylesha and Sravika 2018). It preys on various soft-bodied arthropods (Gupta et al. 2020; Lenin and Rajan 2016; Pan et al. 2020; Vanitha et al. 2018; Yu et al. 2021). The stink bug can feed on more than 40 types of pests as both nymphs and adults and can be readily reared in laboratories or natural enemy insect rearing facilities, making it a valuable natural enemy insect with great potential for use in integrated pest management (Guo et al. 2021; Gupta et al. 2014; Pan et al. 2022; Zhang et al. 2023).

Locomotion is a fundamental behavior regulated by the central nervous system, motor neurons, and muscles in animals. Various motor behaviors such as walking, flying, swimming, and crawling, characterized predominantly by rhythmic movement, are critically important for survival and reproduction (De Jongh 2021; Mantziaris et al. 2020; Orlovsky et al. 1999). Insects move to find food, mating partners, and oviposition sites and avoid predators. Walking activity, which consumes less energy than flying, is the primary form of short-distance movement in insects (Shen et al. 2020). Generally, the wingless nymphs and winged adults of *E. furcellata* travel by walking to find food, mates, and oviposition sites on the same or adjacent plants. Therefore, walking behavior is crucial to the survival and reproduction of these stink bugs. However, the effects of widely used imidacloprid on the walking behavior of stink bugs are still unknown. Herein, we examined how acute exposure to imidacloprid might impair the walking behavior of stink bugs by using a locomotion compensator.

## Materials and Methods

**Insects.** Eocanthecona furcellata adults were originally provided by GreenLeaf Biological Control Company (Yuxi, Yunnan Province) and were reared in nylon cages (45 cm  $\times$  45 cm  $\times$  45 cm) under laboratory conditions at 25  $\pm$  1°C, 50–70% relative humidity, and a photoperiod of 14:10 (L:D) h. The eggs of the stink bug were collected into a plastic box containing a moist sponge; all nymphs were reared in cages after hatching. The nymphs of *Tenebrio molitor* L. were provided for stink bug development. Adults 2–7 d after emergence and nymphs were used in the experiments.

**Chemicals.** Imidacloprid was purchased from Sigma-Aldrich (St. Louis, MO) and diluted with double-distilled water. Field surveys showed that imidacloprid at a dose of 0.04375% (two bags of 70% imidacloprid dissolved in  $\sim$ 16 kg of water) is commonly used to spray in tobacco, cotton, and rice paddy systems. Therefore,

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1/500th of the field-realistic dose of imidacloprid (0.875 ppm) was used in the behavioral experiments.

**Behavioral bioassay.** The walking behavior of the stink bug was tested with a TrackSphere LC-300 locomotion compensator (Syntech, Hilversum, The Netherlands), as described by Piesik et al. (2009). This device is composed of a white sphere (300 mm in diameter) coupled to two servomotors driven by a controller (TrackSphere TRC-01 controller, Syntech). A video camera mounted directly above the sphere monitored the movements of the insect and was connected to the controller that drives the servomotors to compensate for the movements of the insect, so that the image of the insect remained in the center of the visual field. The stink bug's track was recorded by the tracking software TrackSphere<sup>TM</sup> v 3.1 (Syntech) at a rate of 10 Hz.

Stink bugs were submerged in double-distilled water or imidacloprid for 10 s before being removed and used in behavioral tests after 30 min. Fifteen bugs were used in each treatment group. The mean walking speed, walking time, and walking distance were recorded by the tracking software.

**Statistical analysis.** All data are presented as means  $\pm$  SEM, with a threshold level of statistical significance set at *P* < 0.05. An independent sample *t* test was used to analyze the differences in mean walking speed, walking time, and walking distance between control (CTL) and imidacloprid (IMI) groups.

## Results

**Walking speed.** Compared with CTL groups, the mean walking speeds of the IMI groups (centimeters per second, IMI versus CTL) were significantly reduced for first–fifth instar nymphs and adults (first instar:  $0.381 \pm 0.024$  versus  $1.191 \pm 0.048$ , P < 0.001; second instar:  $0.492 \pm 0.035$  versus  $1.012 \pm 0.053$ , P < 0.001; third instar:  $0.560 \pm 0.037$  versus  $1.832 \pm 0.107$ , P < 0.001; fourth instar:  $1.093 \pm 0.067$  versus  $2.107 \pm 0.061$ , P < 0.001; fifth instar:  $1.233 \pm 0.103$  versus  $2.609 \pm 0.140$ , P < 0.001; adult female:  $2.040 \pm 0.099$  versus  $3.920 \pm 0.071$ , P < 0.001; and adult male:  $1.493 \pm 0.085$  versus  $3.685 \pm 0.103$ , P < 0.001) (Fig. 1).

**Walking time.** Likewise, compared with CTL groups, the walking times of the IMI groups (seconds, IMI versus CTL) were significantly decreased for first–fifth instar nymphs and adults (first instar:  $317.9 \pm 22.4$  versus  $1,174.1 \pm 92.0$ , P < 0.001; second instar:  $599.3 \pm 44.4$  versus  $1,643.3 \pm 72.0$ , P < 0.001; third instar:  $709.9 \pm 40.5$  versus  $2,282.5 \pm 155.0$ , P < 0.001; fourth instar:  $671.3 \pm 42.0$  versus  $1,860.1 \pm 71.7$ , P < 0.001; fifth instar:  $906.7 \pm 79.5$  versus  $2,240.1 \pm 73.8$ , P < 0.001; adult female:  $806 \pm 69.5$  versus  $1,673.6 \pm 43.1$ , P < 0.001; and adult male:  $659.4 \pm 63.6$  versus  $1,833.2 \pm 33.5$ , P < 0.001) (Fig. 2).

**Walking distance.** Compared with CTL groups, the walking distances of the IMI groups (IMI vs. CTL, centimeters) were significantly diminished for the first–fifth instar nymphs and adults (first instar:  $11,824.0 \pm 987.1$  versus  $141,346.1 \pm 12,698.5$ , P < 0.001; second instar:  $28,939.2 \pm 2,583.5$  versus  $168,899.6 \pm 15,601.7$ , P < 0.001; third instar:  $38,989.2 \pm 2,737.4$  versus  $432,168.7 \pm 49,602.4$ , P < 0.001; fourth instar:  $73,449.0 \pm 6,431.2$  versus  $390,784.7 \pm 17,479.7$ , P < 0.001; fifth instar:  $116,342.2 \pm 16,840.8$  versus  $585,366.2 \pm 37,418.1$ , P < 0.001; adult female:  $165,481.3 \pm 16,648.6$  versus  $657,636.1 \pm 23,693.1$ , P < 0.001; and adult male:  $101,315.7 \pm 11,277.2$  versus  $676,847.9 \pm 25,114.4$ , P < 0.001) (Fig. 3).

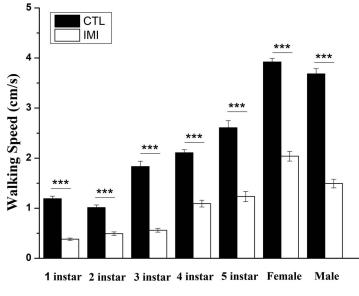


Fig. 1. Reduced walking speed of stink bug.

# Discussion

Our study is the first to test how a commonly used neonicotinoid insecticide directly affects the walking behaviors of natural enemy insects. The major findings of our study demonstrate that acute exposure to imidacloprid significantly affected mean walking speed, walking time, and walking distance in both nymphs and

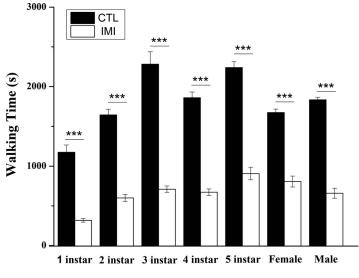


Fig. 2. Decreased walking time of stink bug.

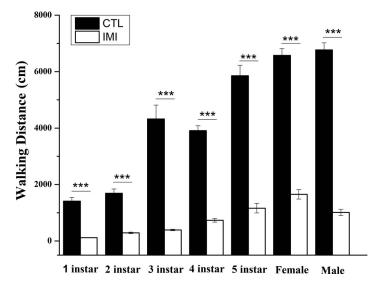


Fig. 3. Decreased walking distance of stink bug.

adults of *E. furcellata* (30.6–52.0% in mean walking speed; 27.1–48.2% in walking time; 8.4–25.2% in walking distance; Figs. 1–3). Together, these results indicate that imidacloprid impairs the walking behavior of *E. furcellata*, suggesting that attention should be paid to the negative effects of imidacloprid when releasing, augmenting, or conserving natural enemy insects in production fields.

It is well known that walking is crucial for insects to move around in search of food, mating partners, oviposition sites, and avoidance of predators. Eocanthecona furcellata can encounter nymphs of lepidopteran and coleopteran pests in the same or adjacent crops after being released into fields. Because of the widespread use of imidacloprid in agricultural fields, exposure to this pesticide is unavoidable for the stink bug. Previous studies have shown that imidacloprid impairs social interaction (Desneux et al. 2007), associative learning (Mengoni Goñalons and Farina 2018; Mustard et al. 2020; Smith et al. 2020), flying behavior (Kenna et al. 2019), and effective foraging and pollination services (Stanley et al. 2015) in honey bees and bumblebees. Imidacloprid affects gustatory sensitivity to sucrose and impairs the ability of honey bees to perform the waggle dance (Eiri and Nieh 2012). Imidacloprid reduces the ability of bees to forage and perform homing flights in field situations (Bortolotti et al. 2003; Mommaerts et al. 2010; Schneider et al. 2012). High acute doses (100 and 500 ppb) of imidacloprid affect locomotion in bees (Medrzycki et al. 2003). A recent study showed that imidacloprid at acute doses of 200 and 400 ppb significantly reduces overall daily activity in honey bees (Delkash-Roudsari et al. 2020), and imidacloprid at a dose of 10 ppb reduces flight time (Kenna et al. 2019), showing similar patterns of effects to our findings.

Motor activity is the output of nerve signals and is the result of the combined action of the central nervous system, motor neurons, and muscles (De Jongh 2021; Mantziaris et al. 2020). Acetylcholine is an important excitatory neurotransmitter in the insect nervous system, regulating the input and output of nerve signals (Gauthier

2010; Hu et al. 2010). Previous studies have shown that imidacloprid has a high agonistic affinity for nicotinic acetylcholine receptors, affecting the central nervous system (Elbert et al. 2008; Schmuck et al. 2003). Imidacloprid binds to acetylcholine receptors, inhibiting or blocking nerve signal transmission, which may lead to reduced movement speed, power, and ultimately, shortened movement distance in the stink bugs. In addition, imidacloprid has been shown to induce the down-regulation of genes involved in sugar-metabolizing pathways (Derecka et al. 2013) and could also reduce mitochondrial activity, impair respiratory processes, and cause rapid mitochondrial depolarization (Moffat et al. 2015; Nicodemo et al. 2014), which may contribute to the impaired walking performance in stink bugs. To understand the neural and molecular mechanisms by which imidacloprid impairs walking behavior, further research is needed to determine where and how imidacloprid affects nerves and muscles during signal transmission.

In summary, our results show that acute exposure to imidacloprid impaired the walking behavior of *E. furcellata*, indicating that we should pay attention to the negative effects of neonicotinoid pesticides on the pest control efficacy of natural enemy insects in the future.

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### **References Cited**

- Bonmatin, J.M., C. Giorio, V. Girolami, D. Goulson, D.P. Kreutzweiser, C. Krupke, M. Liess, E. Long, M. Marzaro, E.A.D. Mitchell, D.A. Noome, N. Simon-Delso and A. Tapparo. 2015. Environmental fate and exposure; neonicotinoids and fipronil. Environ. Sci. Pollut. Res. Int. 22: 35–67.
- Bortolotti, L., R. Montanari, J. Marcelino, P. Medrzycki, S. Maini and C. Porrini. 2003. Effects of sublethal imidacloprid doses on the homing rate and foraging activity of honey bees. Bull. Insectol. 56: 63–67.
- Blacquière, T., G. Smagghe, C.A.M. van Gestel and V. Mommaerts. 2012. Neonicotinoids in bees: A review on concentrations, side-effects and risk assessment. Ecotoxicology 21: 973–92.
- **De Jongh, E. 2021.** Navigation and orientation in Coleoptera: A review of strategies and mechanisms. Anim. Cogn. 24: 1153–1164.
- Delkash-Roudsari, S.A.M. Chicas-Mosier, S.H. Goldansaz, K. Talebi-Jahromi, A. Ashouri and C.I. Abramson. 2020. Assessment of lethal and sublethal effects of imidacloprid, ethion, and glyphosate on aversive conditioning, motility, and lifespan in honey bees (*Apis mellifera* L.). Ecotoxicol. Environ. Saf. 204: 111108.
- Derecka, K., M.J. Blythe, S. Malla, D.P. Genereux, A. Guffanti, P. Pavan, A. Moles, C. Snart, T. Ryder, C.A. Ortori, D.A. Barrett, E. Schuster and R. Stöger. 2013. Transient exposure to low levels of insecticide affects metabolic networks of honeybee larvae. PLoS One 8: e68191.
- Desneux, N., A. Decourtye and J.M. Delpuech. 2007. The sublethal effects of pesticides on beneficial arthropods. Annu. Rev. Entomol. 52: 81–106.
- Eiri, D.M. and J.C. Nieh. 2012. A nicotinic acetylcholine receptor agonist affects honey bee sucrose responsiveness and decreases waggle dancing. J. Exp. Biol. 215(Pt 12): 2022-9.
- Elbert, A., M. Haas, B. Springer, W. Thielert and R. Nauen. 2008. Applied aspects of neonicotinoid uses in crop protection. Pest Manag. Sci. 64: 1099–1105.

- Environmental Protection Agency. 2015. Colony collapse disorder: European bans on neonicotinoid pesticides. 20 February 2015. (http://www.epa.gov/pesticides/about/intheworks/ ccd-european-ban.html)
- Gauthier, M. 2010. State of the art on insect nicotinic acetylcholine receptor function in learning and memory, Pp. 97–115. *In* Thany, S.H. (ed.), Insect Nicotinic Acetylcholine Receptors. Springer, New York, NY.
- Godfray, H., J. Charles, T. Blacquière, R.S. Hails, S.G. Potts, N.E. Raine, A.J. Vanbergen and A.R. McLean. 2015. A restatement of recent advances in the natural science evidence base concerning neonicotinoid insecticides and insect pollinators. Proc. R. Soc. B. 282: 20151821.
- Guo, Y., J.J. Xiao, D.S. Li and J. Wang. 2021. The complete mitochondrial genome of the stink bug *Eocanthecona furcellata* (Hemiptera: Pentatomidae). Mitochondrial DNA B Resour. 6: 3085–3086.
- Gupta, R.K., M. Gani, P. Jasrotia, K. Srivastava and V. Kaul. 2014. A comparison of infectivity between polyhedra of the *Spodoptera litura* multiple nucleopolyhedroviral before and after passage through the gut of the stink bug, *Eocanthecona furcellata*. J. Insect. Sci. 14: 96.
- He, X.N., J.D. Xian, R. Chen, Z.Y. Zhang and R. Zeng. 2013. Effects of four insect feed on development and reproduction of *Cantheconidea furcellata* (Hemiptera: Asopinae). J. Environ. Entomol. 35: 799–803.
- Hu, A., W. Zhang and Z. Wang. 2010. Functional feedback from mushroom bodies to antennal lobes in the Drosophila olfactory pathway. Proc. Natl. Acad. Sci. USA 107: 10262–10267.
- Jeschke, P., R. Nauen, M. Schindler and A. Elbert. 2011. Overview of the status and global strategy for neonicotinoids. J. Agric. Food. Chem. 59: 2897–2908.
- Keerthi, M.C., A. Sravika, H.S. Mahesha, A. Gupta, H.A. Bhargavi and S. Ahmed. 2020. Performance of the native predatory bug, *Eocanthecona furcellata* (Wolff) (Hemiptera: Pentatomidae), on the fall armyworm, *Spodoptera frugiperda* (J.E. Smith) (Lepidoptera: Noctuidae), and its limitation under field condition. Egypt. J. Biol. Pest Control 30: 69.
- Kenna, D., H. Cooley, I. Pretelli, A. Ramos Rodrigues, S.D. Gill and R.J. Gill. 2019. Pesticide exposure affects flight dynamics and reduces flight endurance in bumblebees. Ecol. Evol. 9: 5637–5650.
- Lenin, E.A. and S.J. Rajan. 2016. Biology of predatory bug *Eocanthecona furcellata* (Wolff) (Hemiptera: Pentatomidae) on *Corcyra cephalonica* Stainton. J. Entomol. Zool. Stud. 4: 338–340.
- Mantziaris, C., T. Bockemühl and A. Büschges. 2020. Central pattern generating networks in insect locomotion. Dev. Neurobiol. 80: 16–30.
- Matsuda, K., S.D. Buckingham, D. Kleier, J.J. Rauh, M. Grauso and D.B. Sattelle. 2001. Neonicotinoids: Insecticides acting on insect nicotinic acetylcholine receptors. Trends Pharmacol. Sci. 22: 573–580.
- Medrzycki, P., R. Montanari, P. Bortolotti, A. Sabatini, S. Maini and C. Porrini. 2003. Effects of imidacloprid administered in sublethal doses on honey bee behaviour. Laboratory tests. Bull. Insectol. 56: 59–62.
- Mengoni Goñalons, C. and W.M. Farina. 2018. Impaired associative learning after chronic exposure to pesticides in young adult honey bees. J. Exp. Biol. 221(Pt 7): jeb176644.
- Mitchell, E.A.D., B. Mulhauser, M. Mulot, A. Mutabazi, G. Glauser and A. Aebi. 2017. A worldwide survey of neonicotinoids in honey. Science 358: 109–111.
- Moffat, C., J.G. Pacheco, S. Sharp, A.J. Samson, K.A. Bollan, J. Huang, S.T. Buckland and C.N. Connolly. 2015. Chronic exposure to neonicotinoids increases neuronal vulnerability to mitochondrial dysfunction in the bumblebee (*Bombus terrestris*). FASEB J. 29: 2112–2119.
- Mommaerts, V., S. Reynders, J. Boulet, L. Besard, G. Sterk and G. Smagghe. 2010. Risk assessment for side-effects of neonicotinoids against bumblebees with and without impairing foraging behaviour. Ecotoxicology 19: 207–215.

- Mustard, J.A., A. Gott, J. Scott, N.L. Chavarria and G.A. Wright. 2020. Honeybees fail to discriminate floral scents in a complex learning task after consuming a neonicotinoid pesticide. J. Exp. Biol. 223(Pt 5): jeb217174.
- Nicodemo, D., M.A. Maioli, H.C.D. Medeiros, M. Guelfi, K.V. Balieira, D. De Jong and F.E. Mingatto. 2014. Fipronil and imidacloprid reduce honeybee mitochondrial activity. Environ. Toxicol. Chem. 33: 2070–2075.
- **Orlovsky, G.N., T.G. Deliagina and S. Grillner. 1999.** Neuronal control of locomotion: From mollusc to man. Oxford Univ. Press, Oxford, U.K.
- Pan, Y.N., M.Y. Yao, D. Fu, F. Guo, R. Xun, L. Jiang and W.L. Chen. 2020. Predation effects of Eocanthecona furcellata (Wolff) adults against *Spodoptera frugiperda* nymphs. J. Mt. Agric. Biol. 39: 23–27.
- Pan, Y.N., R.N. Zhao, D. Fu, C. Yu, C.N. Pan, W. Zhou and W.L. Chen. 2022. Assessment of suitable reference genes for qRT-PCR normalization in *Eocanthecona furcellata* (Wolff). Insects 13: 773.
- Piesik, D., D. Rochat, J. van der Pers and F. Marion-Poll. 2009. Pulsed odors from maize or spinach elicit orientation in European corn borer neonate larvae. J. Chem. Ecol. 35: 1032–1042.
- Rider, D. and L. Zheng. 2002. Checklist and nomenclatural notes on the Chinese Pentatomidae (Heteroptera). I. Asopinae. Entomotaxonomia 24: 107–115.
- Robin, S. and S. Andreas. 2003. Uptake, translocation and metabolism of imidacloprid in plants. Bull. Insectol. 56: 35–40.
- Roca-Cusachs, M., J. Kim, J. Park and S. Jun. 2020. Taxonomic review of the predatory stink bugs of the Korean Peninsula (Heteroptera: Pentatomidae: Asopinae), with a key to the Korean species and a discussion of their usefulness as biological control agents. J. Asia Pac. Entomol. 23: 113–123.
- Schmuck, R., R. Nauen and U. Ebbinghaus-Kintscher. 2003. Effects of imidacloprid and common plant metabolites of imidacloprid in the honey bee: toxicological and biochemical considerations. Bull. Insectol. 56: 27–34.
- Schneider, C.W., J. Tautz, B. Grünewald and S. Fuchs. 2012. RFID tracking of sublethal effects of two neonicotinoid insecticides on the foraging behavior of *Apis mellifera*. PLoS One 7: e30023.
- Shen, Z.Y., You, X.F., Zhang, S.H., C.N. Zhang, X.J. Yang, H.S. Xing, W.Z. Li and G.H. Yuan. 2020. Principle component analysis on walking parameters of *Holotrichia parallela* beetles obtained from Tracksphere locomotion compensator. Entomol. Central China 16: 257–266.
- Smith, D.B., A.N. Arce, A. Ramos Rodrigues, P.H. Bischoff, D. Burris, F. Ahmed and R.J. Gill. 2020. Insecticide exposure during brood or early-adult development reduces brain growth and impairs adult learning in bumblebees. Proc. Biol. Sci. 287: 20192442.
- Simon-Delso, N., V. Amaral-Rogers, L.P. Belzunces, J.M. Bonmatin, M. Chagnon, C. Downs, L. Furlan, D.W. Gibbons, C. Giorio, V. Girolami, D. Goulson, D.P. Kreutzweiser, C.H. Krupke, M. Liess, E. Long, M. McField, P. Mineau, E.A. Mitchell, C.A. Morrissey, D.A. Noome, L. Pisa, J. Settele, J.D. Stark, A. Tapparo, H. Van Dyck, J. Van Praagh, J.P. Van der Sluijs, P.R. Whitehorn and M. Wiemers. 2015. Systemic insecticides (neonicotinoids and fipronil): Trends, uses, mode of action and metabolites. Environ. Sci. Pollut. Res. Int. 22: 5–34.
- Stanley, D.A., M.P. Garratt, J.B. Wickens, V.J. Wickens, S.G. Potts and N.E. Raine. 2015. Neonicotinoid pesticide exposure impairs crop pollination services provided by bumblebees. Nature 528: 548–550.
- Shylesha, A.N. and A. Sravika. 2018. Natural occurrence of predatory bugs, *Eocanthecona furcellata* (Wolff) and *Andrallus spinidens* (Fabr.) on *Spodoptera frugiperda* (Smith) (Hemiptera: Pentatomidae) in maize and their potential in management of fall armyworm. J. Biol. Control 32: 209–211.

- Vanitha, K., T.N. Raviprasad and V.R. Shwetha. 2018. Life cycle of *Eocanthecona furcellata* Wolff. (Hemiptera: Pentatomidae) a predatory bug in cashew plantations, upon rearing on wax moth larvae. J. Entomol. Zool. Stud. 6: 3007–3010.
- Yu, C., M.Y. Yao, R.N. Zhao, L. Wang and W.L. Chen. 2021. Effects of breeding density on growth and development of *Eocanthecona furcellata* (Wolff). J. Mt. Agric. Biol. 40: 75–79.
- Zhang, M., Z. Dai, X. Chen, D. Qin, G. Zhu, T. Zhu, G. Chen, Y. Ding, G. Wu and X. Gao. 2023. Identification and functional analysis of serine protease inhibitor gene of *Eocanthecona furcellata* (Wolff). Front. Physiol. 14: 1248354.